# Enhancement of NSIC Rc 192 Seedling Growth by Soil-based and Carbonized Rice Hull (CRH)-based Actinomycete Inoculants

Kassandra Jin B. Mariano<sup>1</sup>, Ann Jhudeil C. Santos<sup>2</sup>, Editha V. Evangelista<sup>2</sup> and Jayvee A. Cruz<sup>2\*</sup> <sup>1</sup>Department of Biological Sciences Central Luzon State University Science City of Muñoz, Nueva Ecija 3120 Philippines

<sup>2</sup>Agronomy, Soils, and Plant Physiology Division Philippine Rice Research Institute Science City of Muñoz, Nueva Ecija 3120 Philippines *\*jayveecruz@gmail.com* 

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#### Abstract

A study was conducted to observe the effects of soil-based (SB) and carbonized rice hull (CRH)-based actinomycete inoculants on the seedling growth of rice variety NSIC Rc 192. The experiment was conducted under laboratory conditions using wet paper towels in petri dishes. The statistical design of the experiment was completely randomized (CRD) with three replicates per treatment. The actinomycete isolate, Streptomyces sp., used in the study was previously reported to produce indole-3-acetic acid (IAA), 1-aminocyclopropane-1-carboxylate (ACC) deaminase, and phosphatase. In this experiment, rice seeds were treated with soil-based and CRH-based actinomycete inoculants. Growth parameters such as shoot and root length and oven dry weight were measured 7 days after sowing (DAS). Inoculation with CRH-based and soil-based actinomycete inoculants significantly increased shoot length by 102.61% and 94.77%, respectively, relative to the uninoculated treatment at 7 DAS. Inoculation with CRH-based and soil-based actinomycete inoculants significantly increased root length by 113.24% and 98.53%, respectively, relative to the uninoculated treatment at 7 DAS. The highest shoot (4.5 mg) and root (3.5 mg) oven dry weight was observed at CRH-based inoculation while the lowest shoot (2.0 mg) and root (0.5 mg) was obtained at the uninoculated control. Regardless of the carrier used, actinomycete isolate can enhance the growth of rice seedlings. Both CRH and soil-based actinomycete inoculants significantly increased the shoot and root length and oven dry weight of rice seedlings.

*Keywords*: actinomycetes, inoculant carrier, plant growth-promoting bacteria (PGPB), rice, seed germination

## 1. Introduction

The actinomycetes are the group of most filamentous bacteria forming long filaments that stretch through the soil. They are prokaryotic, gram-positive, and most are mesophilic. The growth of actinomycetes is inhibited at pH 5.0 or higher and when the moisture content reaches 85-100% or flooding occurs (Prigent, 2012). Actinobacteria can solubilize phosphate, promote plant growth, and produce siderophore and phytohormone (IAA) (Jog et al. 2014). The use of plant growth-promoting bacteria, specifically actinomycetes, has been gaining the attention of the researchers in the past few years (Ahemad and Kibret, 2014). Streptomyces, a genus under actinomycete, is one of the many widely studied genera receiving increasing popularity because of its plant growth-promoting properties (Sousa and Olivares, 2016). Species under Streptomyces are well-known to be isolated from the soil around the roots which raises many concerns on the significance of the bacteria to its host (Sousa and Olivares, 2016). Streptomyces species have been isolated from different environments and was reported to exhibit a wide range of plant growth-promoting activities which include its ability to produce IAA, ACC deaminase, siderophores, and its phosphate-solubilizing activity (Sadeghi et al., 2012; Abd-Alla et al., 2013).

Several studies have reported the effectiveness of Streptomyces in improving and enhancing the growth and yield of different crops. In the study of Gopalakrishnan et al. (2013), Streptomyces inoculation significantly enhanced all the agronomic traits of sorghum and rice under greenhouse and field conditions, respectively. A similar study of Gopalakrishnan et al. (2015) also demonstrated the effectiveness of the same Streptomyces strains in enhancing the growth parameters, grain yield, and total dry matter of chickpea plant. However, information about the effect of the actinomycete during the germination of rice was not provided in these studies. One of the few studies that provided information on the impact of the actinomycete during the germination of rice showed that a siderophore-producing endophytic streptomycete isolated from roots of a Thai jasmine rice plant enhanced the growth of rice and mungbean plants (Rungin et al., 2012). The rate of rice growth during the early stage, especially after germination, is very critical since it will determine the success of crop establishment (Ogiwara and Terashima, 2001). Improving the plant growth and vigor during this stage is also essential for robustness and survival of the seedlings under adverse conditions (Vibhuti et al., 2015). Therefore, inoculation of actinomycete with an ability to produce ACC deaminase, IAA and phosphatase during early establishment of rice seedlings is beneficial. Hence, the objectives of this.

# 2. Methodology

## 2.1 Time and Place of the Study

The experiment was conducted under laboratory conditions from June to July 2017 at the Philippine Rice Research Institute - Central Experiment Station (PhilRice-CES), Science City of Muñoz, Nueva Ecija, Philippines (15° 40' N, 120° 53' E, 57.6 masl). Experimental units were placed under laboratory temperature ( $28 \pm 2$  °C) with alternating light-dark cycles. Each of the three treatments in the study was done in triplicates.

## 2.2 Isolate Used

Actinomycete was isolated from Binangonan soil in Rizal, Philippines. This bacterium has been previously proven to produce plant growth-promoting compounds such as ACC deaminase, IAA, and phosphatase, which effectively promoted rice growth under laboratory room conditions (Cruz *et al.*, 2014; Cruz *et al.*, 2015c). The probable actinomycete identity is *Streptomyces mutabilis* with 98% of maximum identity based on 16S rDNA analysis (Cruz *et al.*, 2015d). The bacterium was maintained on arginine-glycerol-salt (AGS) agar slants.

## 2.3 Seed Surface Sterilization

NSIC Rc 192 seeds were washed with tap water for five times to eliminate unwanted particles then soaked in 95% ethyl alcohol for 2 ½ minutes and washed with sterile distilled water five times. The seeds were then soaked in 30% sodium hypochlorite (NaClO) for 30 seconds and washed again with sterile distilled water five times to rinse off the 30% NaClO.

## 2.4 Preparation Soil-based and CRH-based Carriers

For the preparation of soil-based carrier, components (soil and charcoal) were pulverized, sieved, and mixed, following the ratio of 3 soil: 1 charcoal. One hundred grams (g) of the mixture was weighed and packed into autoclavable plastic bags and sterilized for 1 hour at 121 °C for three consecutive days.

For the preparation of CRH-based carrier, 100 g of CRH was weighed and packed into autoclavable plastic bags and sterilized for one hour at 121 °C for three consecutive days.

#### 2.5 Preparation of Soil-based and CRH-based Inoculants

A loopful of actively growing *Streptomyces* sp. was inoculated into 50 mL and 100 mL of Arginine Glycerol Salt (AGS) broth and was incubated for five to seven days at room temperature (28-30 °C). After incubation, 50 mL and 100 mL of the culture broths were aseptically inoculated into the 100g sterilized soil-based and 100g CRH-based carriers, respectively, which brought the soil and CRH moisture to approximately field capacity.

## 2.6 Inoculant Suspension Preparation and Application

Five grams of each of the actinomycete inoculant were added to a separate 100 ml sterile distilled water in a beaker. The surface sterilized seeds were separately soaked in each inoculant suspension for 30 minutes.

#### 2.7 Seedling of NSIC Rc192

CRH and SB inoculant suspensions were used to moist the previously sterilized paper towels in Petri plates. Twenty CRH-based and SB actinomycete inoculant-treated seeds were sown into the plates using sterile forceps. Seeds soaked in sterile distilled water served as the control. The experimental set-up was watered using sterile distilled water for seven days to maintain the moist environment.

#### 2.8 Gathering of Data

Shoot and root length (cm) of five randomly sampled seedlings from each treatment was measured seven DAS (Agbodjato *et al.*, 2016; Gangwar, 2013; Gopalakrishnan *et al.*, 2013). The total oven dry weight was also recorded after 72 hours of oven-drying.

#### 2.9 Statistical Analysis

Statistical analysis was performed using statistical analysis system (SAS) portable v.9.0. All data gathered were analyzed using one way analysis of variance (ANOVA) in a completely randomized design (CRD). Significant differences between treatments were determined using least significant difference (LSD) test at p<0.05.

## 3. Results and Discussion

Table 1 shows the growth of rice as affected by actinomycete inoculation at seven DAS. Both treatments which include inoculation of *Streptomyces* sp. significantly increased the shoot and root length of the germinated rice relative to the uninoculated control (Figure 1).

Table 1. The growth of NSIC Rc 192 as affected by CRH-based and soil-based actinomycete inoculants at seven DAS

Treatment		Shoot length (cm)	Root length (cm)	Shoot oven dry weight (mg/3 seedlings)	Root oven dry weight (mg/3 seedlings)
1. 2.	Uninoculated CRH-based	3.06 <sup>b</sup>	2.72 <sup>b</sup>	2.00 <sup>c</sup>	0.50°
2.	actinomycete inoculant	6.20 <sup>a</sup>	5.80 <sup>a</sup>	4.00ª	3.50 <sup>a</sup>
3.	SB actinomycete inoculant	5.96ª	5.40 <sup>ab</sup>	3.30 <sup>b</sup>	2.80 <sup>b</sup>

\*Values represent mean of five replications for length and three replications for dry weight. Values with the same letter within a column are not significantly different at P<0.05 according to LSD.



Figure 1. NSIC Rc 192 rice seedlings as affected by actinomycete inoculation: (a) uninoculated control, (b) CRH-based actinomycete inoculant and (c) soil-based actinomycete inoculant

Highest shoot length (6.20 cm) was obtained at rice seedlings treated with carbonized rice hull-based actinomycete inoculant while lowest (3.06 cm) was obtained at the uninoculated treatment. Biocharcoals, such as CRH, are rich source of carbon and are often used as carrier materials (Gaskin *et al.*, 2008). Biocharcoal particles are stable, tiny in size (Chidumayo 1994), and increase the carbon content and fertility of the soil (Steinbeiss *et al.*, 2009). Related studies by Cruz *et al.* (2015a) showed that initial population of actinomycete in CRH carrier increased by 2.2 x 104 cfu/g to 2.9 x 107 cfu/g (135, 169%) five days after inoculation (DAI). Survival of inoculum in a carrier is one of

the major considerations in inoculant production. In this study, inoculation with CRH-based and soil-based actinomycete inoculants significantly increased shoot length by 102.61% and 94.77%, respectively, relative to the uninoculated control at seven DAS.

Highest (5.80 cm) root length was obtained due to CRH-based actinomycete inoculation. On the other hand, the lowest (2.72 cm) root length was obtained at the uninoculated treatment. Inoculation with CRH-based and soil-based actinomycete inoculants significantly increased root length by 113.24% and 98.53%, respectively, relative to the uninoculated control at seven DAS. In a study conducted by Saranya *et al.* (2011), biochar as inoculant carrier significantly increased the root growth, shoot growth, and yield of maize as compared to uninoculated control. One of the reasons for the success of microbial inoculation concerning plant growth promotion can be attributed to the number of viable cells available in the carrier material (Duquenne *et al.*, 1999). On the other hand, a study conducted by Gaind and Gaur (1990) showed that the soil-charcoal mixture is a suitable carrier material in terms of the multiplication of phosphate-solubilizing bacteria that may be due to its higher porosity as compared with the other tested carriers.

Regarding dry weight, both treatments with actinomycete inoculation significantly increased the oven-dried shoot and root weight relative to the uninoculated control. The highest shoot (4.5 mg) and root (3.5 mg) oven-dry weight were obtained due to CRH-based inoculant. The lowest shoot (2.0 mg) and root (0.5 mg) were obtained at the uninoculated control. Inoculation with CRH-based and soil-based actinomycete inoculants significantly increased the shoot oven dry weight by 100% and 65% respectively, relative to the uninoculated treatment at seven DAS. Similarly, inoculation of actinomycete with CRH-based and soil-based actinomycete inoculants significantly increased the root oven dry weight by 600% and 460% respectively, relative to the uninoculated treatment at seven DAS. Related studies by Suralta *et al.* (2017) shows that seminal root length of rice seedlings was 145% greater in actinomycete-inoculated rice seeds than in uninoculated ones at seven days after germination. It suggests that inoculation with actinomycete promotes shoot and root growth of rice seedlings during germination stage.

Related studies reported that *Streptomyces* sp. can enhance the growth of different plants including rice, sorghum, wheat, corn, cucumber, tomato, etc. Gopalakrishnan *et al.* (2013) observed that *Streptomyces* sp. strains significantly enhanced all PGP parameters including root length, volume and

dry weight, and yield parameters over the uninoculated control in both sorghum and rice under greenhouse and field conditions, respectively. The probable reasons for the enhancement of morphological parameters on both *Streptomyces* sp.-treated sorghum and rice could be the ability of the bacteria used to produce IAA and siderophore and or chitinase, lipase, and  $\beta$ -1,3-glucanase.

A study conducted by Hanapi *et al.* (2014) provided positive results on the use of bacteria to enhance the growth of two varieties of rice. *Nitrosomonas europaea* showed better performance on root and shoot length when combined *Rhodopseudomonas palustris* and *Acinetobacter* sp., respectively. Similarly, laboratory experiments conducted by Gangwar (2013) used *P. fluorescens*, a rhizobacterium, as a bioagent on the growth of rice plant and the results showed that the root and shoot length of the rice plant inoculated with the bacteria was significantly higher than the chemical treatment under laboratory conditions. The same results were obtained from the glasshouse conditions; aside from the increased root and shoot length, *P. fluorescens* exhibited superiority compared to the chemically treated rice plants in terms of the increment of fresh and dry root and shoot weight.

In terms of inoculant formulation, scientists are now using different carriers. The soil is usually used, but other materials like CRH can also be a potential inoculant carrier. CRH is a waste product in rice farming, hence is more practical and environment-friendly to use.

The CRH-based and soil-based carriers used in this study had a pH of 7.85 and 6.6, respectively. In a study by Tang *et al.* (2003), actinomycetes were able to survive within the pH range of 6.0-10.0. In terms of the physiological mechanism, cell organelles in the cytoplasm of most organisms are neutral. Enzymes work best at pH close to that of the environment.

In this study, significant differences in terms of effectiveness to rice seedling growth were observed between the two microbial inoculant carriers (soilbased and CRH-based carriers) relative to the uninoculated control. Both CRH and soil-based actinomycete inoculants significantly increased the shoot and root length and oven dry weight of rice seedlings. A previous study on survival test of actinomycete proved that both soil and carbonized rice hull are potential microbial inoculant carriers (Cruz *et al.*, 2015a and Cruz *et al.*, 2015b).

#### 4. Conclusion and Recommendation

Inoculation with CRH-based and soil-based actinomycete inoculants enhanced the growth of rice seedlings. This finding supports that the actinomycete, *Streptomyces* sp., used in the study can improve the rice seedling growth in addition to the other studies which showed similar results. However, it is recommended that further study should be made on the promising actinomycete to firmly establish its effectiveness under field conditions where environmental factors cannot be controlled.

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#### 6. References

Abd-Alla, M.H., El-Sayed, E-S.A., & Rasmey A-H.M. (2013). Indole-3-acetic acid (IAA) production by Streptomyces atrovirens isolated from rhizospheric soil in Egypt. Journal of Biology and Earth Sciences, 3(2), B182-B93.

Agbodjato, N.A., Noumavo, P.A., Adjanohoun, A., Agbessi, L., & Baba-Moussa, L. (2016). Synergistic Effects of Plant Growth Promoting Rhizobacteria and Chitosan on In Vitro Seeds Germination, Greenhouse Growth, and Nutrient Uptake of Maize (Zea mays L.). Biotechnology Research International, 2016, 1-11.

Ahemad, M., & Kibret M. (2014). Mechanisms and applications of plant growth promoting rhizobacteria: current perspective. Journal of King Saud University, 26(1), 1-20.

Chidumayo, E.N. (1994). Effects of wood carbonization on soil and initial development of seedlings in miombo woodland, Zambia. Plant and Soil, 70, 353-357 Cruz, J.A., & Paterno, E.S. (2014). Isolation and Screening of Actinomycetes for its Growth Promoting Activities. Asia Life Sciences (ASL), 23(2), 413-428.

Cruz, J.A., Lantican, N.B., Delfin, E.F., & Paterno, E.S. (2015d). Characterization and Identification of Growth-Promoting Actinomycetes: a Potential Microbial Inoculant. Asia Life Sciences (ASL), 24(1): 383-397.

Cruz J.A., Delfin E.F., & Paterno, E.S. (2015c). Promotion of Upland Rice Growth by Actinomycetes under Growth Room Conditions. Asia Life Sciences (ASL), 24(1), 87-94.

Cruz, J.A., & Cadiente, M.K.M. (2015b). Survival of Actinomycete in a Soil-based Carrier: A Potential Microbial Inoculant. Asia Life Sciences, 24(1), 343-347.

Cruz, J.A., & Cadiente, M.K.M. (2015a). Survival of an Actinomycete in a Carbonized Rice Hull-Based Carrier. IAMURE International Journal of Ecology and Conservation, 15(1). Retrieved from http://ejournals.ph/form/ciid=9338

Duquenne, P., Chenu, C., Richard, G., & Catroux, G. (1999). Effect of carbon source supply and its location on competion between inoculated and established bacterial strains in sterile soil microcosm. FEMS Microbiology Ecology, 29(4), 331-339.

Gangwar, G.P. (2013). Growth promotion of rice seedlings by fungal and bacterial bioagents effective against bacterial leaf blight of rice. Journal of Applied and Natural Science, 5(2), 430-434.

Gaskin, J.W., Steiner, C., Harris, K., Das, K.C., & Bibens B. (2008). Effect of low temperature pyrolysis conditions on biochar for agricultural use. American Society of Agricultural and Biological Engineers, 51, 2061-2069.

Gaind, S., & Gaur A.C. (1990). Shelf life of phosphate-solubilizing inoculants as influenced by type of carrier, high temperature, and low moisture. Canadian Journal of Microbiology, 36, 846-849.

Gopalakrishnan, S., Srinivas, V., Alekhya, G., Prakash, B., Kudapa, H., Rathore, A., & Varshney, R.K. (2015). The extent of grain yield and plant growth enhancement by plant growth-promoting broad-spectrum Streptomyces sp. in chickpea. SpringerPlus, 4(31), 1-10.

Gopalakrishnan, S., Srinivas, V., Vidya, M.S., & Rathore, A. (2013). Plant growthpromoting activities of Streptomyces spp. in sorghum and rice. SpringerPlus, 2(574), 1-8.

Hanapi, S.Z., Supari, N., Alam, S.A.Z., Javed, M.A., Din, A.R.J.M., Tin, L.C., Rashid, S.N.A.A., Annuar, N.A.S., & Sarmidi, M.R. (2014). Microbial Effects on Seed Germination in Malaysian Rice (Oryza sativa L.). Proceedings of the Asia-Pacific Advanced Network, 37, 42-51.

Jog, R., Pandya, M., Nareshkumar, G., & Rajkumar, S. (2014). Mechanism of phosphate solubilization and antifungal activity of Streptomyces spp. isolated from wheat roots and rhizosphere and their application in improving plant growth. Microbiology, 160, 778–788.

Ogiwara, H., & Terashima, K. (2001). A varietal difference in coleoptile growth is correlated with seedling establishment of direct seeded rice in submerged field under low-temperature conditions. Plant Production Science, 4(3), 166-172.

Prigent, A. (2012). Actinomycetes in soil fertility and agricultural production. Retrieved from http://www.akimoo.com/actinomyctes-in-soil-fertility-and-agricultural-production/

Rungin, S., Indananda C., Suttiviriya, P., Kruasuwan, W., Jaemsaeng, R., & Thamchaipenet, A. (2012). Plant growth enhancing effects by a siderophore producing endophytic streptomycete isolated from a Thai jasmine rice plant (Oryza sativa L. cv. KDML105). Antonie Leeuwenhoek, 102, 463-472.

Sadeghi, A., Karimi, E., Dahaji, P.A., Javid, M.G., Dalvand, Y., & Askari, H. (2012). Plant growth promoting activity of an auxin and siderophore producing isolate of Streptomyces under saline soil conditions. World Journal of Microbiology and Biotechnology, 28(4), 1503-1509.

Saranya, K., Krishnan, P.S., Kumutha, K., & French, L. (2011). Potential for Biochar as an Alternate Carrier to Lignite for the Preparation of Biofertilizers in India. International Journal of Agriculture, Environment & Biotechnology, 4(2), 167-172.

Sousa, J.A., & Olivares, F.L. (2016). Plant growth promotion by streptomycetes: ecophysiology, mechanisms and applications. Chemical and Biological Technologies in Agriculture, 3(24).

Steinbeiss, S., Gleixner G., & Antonietti, M. (2009). Effect of biochar amendment on soil carbon balance and soil microbial activity. Soil Biology and Biochemistry, 41, 1301-1310.

Suralta, R.R., Batungbakal, M.Y.T., Bello, J.C.T., Caparas, L.M., Lagunilla, V.H., Lucas, K.M.D., Patungan, J.U., Siping, A.J.O., Cruz, J.A., Cabral, M.C.J., & Niones, J.M. (2017). An Enhanced Root System Developmental Responses to Drought by Inoculation of Rhizobacteria (Streptomyces mutabilis) Contributed to the Improvement of Growth in Rice. Philippine Journal of Science, 147(1), 115-124.

Tang, S.K., Li, W.J., Dong, W., Zhang, Y.G., & Xu, L.H. (2003). Studies of the biological characteristics of some halophilic and halotolerant actinomycetes isolated from saline and alkaline soils. Actinomycetologica, 17, 6-10.

Vibhuti, C.S, Bargali, K., & Bargali, S.S. (2015). Seed germination and seedling growth parameters of rice (Oryza sativa) varieties as affected by salt and water stress. Indian Journal of Agricultural Sciences, 85(1), 102-8.